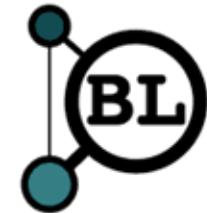


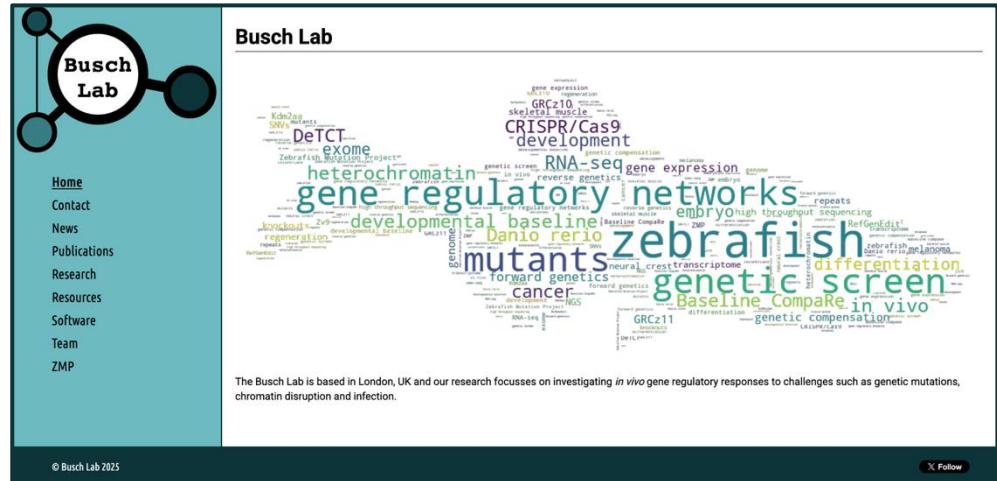
# Introduction to RNA-seq and functional interpretation: Next steps in gene prioritisation

12th Feb 2026



# Me

- Ian Sealy
- Anderson Lab, Sanger Institute
- Previously in Busch Lab, QMUL
- RNA-seq / zebrafish
- Run “*Bioinformatics & Functional Genomics in Zebrafish*” course at EBI



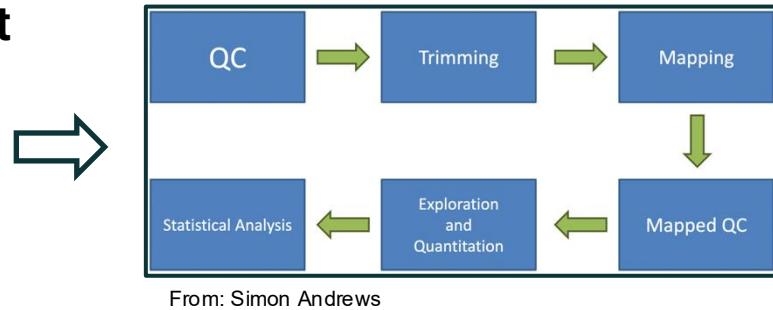
# Questions

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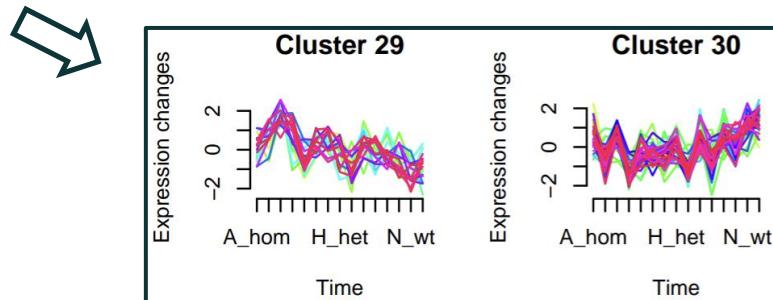
- For timely questions, just unmute and ask
- Or add your question to the Q&A document and I'll answer in a break or later

# Gene list of interest

- Starting point for today: **gene list of interest**
- Most likely from RNA-seq differential expression analysis
- But could be a list from any other analysis:
  - Clustering genes with similar expression profiles
  - Microarray analysis
  - Quantitative proteomics
  - Differential methylation analysis
  - etc...



From: Simon Andrews



# Unranked or ranked gene list?

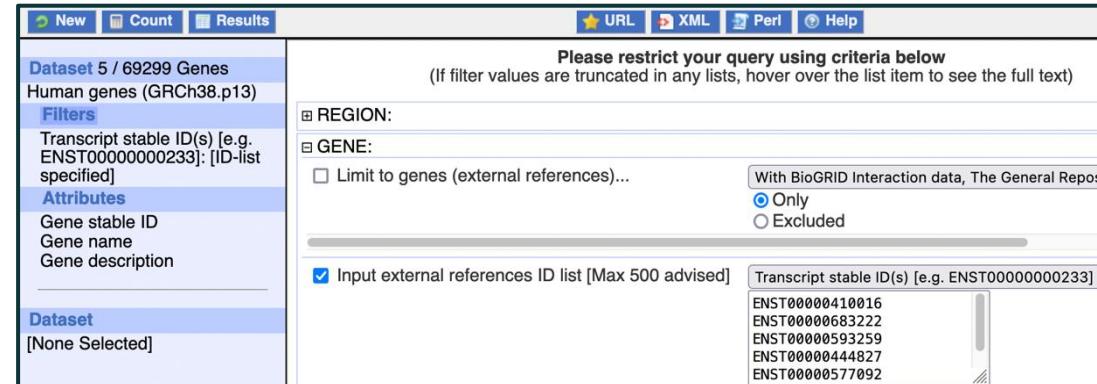
- Gene list can be:
  - Unranked (e.g. genes with somatic mutations in cancer sample)
  - Ranked (e.g. sensitivity in a CRISPR screen)
- RNA-seq differential expression analysis produces ranked lists
- Ranked lists are ordered by a score or metric:
  - e.g. adjusted p-value
  - e.g.  $\log_2$  fold change
- Ranked lists can also have a threshold applied:
  - e.g. adjusted p-value < 0.05

```
ENSDARG00000043198
ENSDARG00000075229
ENSDARG00000036695
ENSDARG00000092115
ENSDARG00000013076
ENSDARG00000015890
ENSDARG00000060682
ENSDARG00000076241
ENSDARG00000093347
ENSDARG00000098114
```

```
ENSDARG00000075676 0.039
ENSDARG00000104197 0.041
ENSDARG00000004301 0.041
ENSDARG00000079766 0.042
ENSDARG00000030494 0.042
ENSDARG00000116804 0.043
ENSDARG00000100599 0.043
ENSDARG00000104325 0.043
ENSDARG00000111102 0.043
ENSDARG00000022466 0.044
```

# “Gene” list of interest

- May not actually be a list of genes
- Could be transcripts or proteins or SNPs, etc...
- Most tools require a list of genes so need to convert
- BioMart is a useful tool for conversions (and other bioinformatics tasks):  
[www.ensembl.org/biomart/martview](http://www.ensembl.org/biomart/martview)



The screenshot shows the BioMart interface. At the top, there are buttons for 'New', 'Count', and 'Results'. On the right, there are links for 'URL', 'XML', 'Perl', and 'Help'. The main area has a header: 'Please restrict your query using criteria below' and '(If filter values are truncated in any lists, hover over the list item to see the full text)'. On the left, there are sections for 'Filters' and 'Attributes'. The 'Filters' section contains a 'REGION' checkbox and a 'GENE' checkbox. Under 'GENE', there is a 'Limit to genes (external references)...' checkbox, a radio button for 'Only' (selected), and another for 'Excluded'. The 'Attributes' section lists 'Gene stable ID', 'Gene name', and 'Gene description'. The 'Dataset' section shows 'Dataset 5 / 69299 Genes' and 'Human genes (GRCh38.p13)'. At the bottom, there is a list of transcript stable IDs: ENST00000410016, ENST00000683222, ENST00000593259, ENST00000444827, and ENST00000577092.

# What next?

- Have a gene list, but what do you do next?
- How do you relate the gene list to existing knowledge?

Gene	pval	adjp	log2fc
ENSDARG00000041294	4.904002310063973e-37	1.0867269119101765e-32	1.5709251030700861
ENSDARG00000060498	1.1297090308658515e-25	1.2517176061993635e-21	1.5921762041345
ENSDARG00000031683	3.2009883731403506e-25	2.364463411626339e-21	-1.277820860357806
ENSDARG00000077982	5.3336179195843655e-18	2.9548243274497384e-14	0.9349522690823255
ENSDARG00000070480	1.2940060161760502e-17	5.735034663692255e-14	1.0699010828953783
ENSDARG0000007769	4.245003753873642e-17	1.5678213864306653e-13	1.6785196633873156
ENSDARG00000102435	6.025610180317608e-17	1.9075360227976884e-13	1.0539265022132713
ENSDARG00000101482	9.742460938723084e-17	2.6986616800262944e-13	0.9350743176658163
ENSDARG0000034503	2.261103100242347e-16	5.567338300152267e-13	0.6082489350504545

# What next?

- Have a gene list, but what do you do next?
- How do you relate the gene list to existing knowledge?
- Add annotation (e.g. BioMart)

Gene	pval	adjp	log2fc
ENSDARG00000041294	4.904002310063973e-37	1.0867269119101765e-32	1.5709251030700861

Gene	pval	adjp	log2fc	Chr	Start	End	Name	Description
ENSDARG00000041294	4.904002310063973e-37	1.0867269119101765e-32	1.5709251030700861	3	62161184	62169060	noxo1a	NADPH oxidase organizer 1a
ENSDARG00000060498	1.1297090308658515e-25	1.2517176061993635e-21	1.5921762041345	23	30006206	30010042	tnfrsf9a	tumor necrosis factor receptor superfamily, member 9a
ENSDARG00000031683	3.2009883731403506e-25	2.364463411626339e-21	-1.277820860357806	20	46552311	46554440	fosab	v-fos FBJ murine osteosarcoma viral oncogene homolog Ab
ENSDARG00000077982	5.3336179195843655e-18	2.9548243274497384e-14	0.9349522690823255	22	661505	665371	elf3	E74-like factor 3 (ets domain transcription factor, epithelial-specific)
ENSDARG00000070480	1.2940060161760502e-17	5.735034663692255e-14	1.0699010828953783	19	30400372	30404096	agr2	anterior gradient 2
ENSDARG0000007769	4.245003753873642e-17	1.5678213864306653e-13	1.6785196633873156	7	56602521	56606752	sult5a1	sulfotransferase family 5A, member 1
ENSDARG00000102435	6.025610180317608e-17	1.9075360227976884e-13	1.0539265022132713	7	45975537	45976956	plekhf1	pleckstrin homology domain containing, family F (with FYVE domain) member 1
ENSDARG00000101482	9.742460938723084e-17	2.6986616800262944e-13	0.935074317658163	5	13870340	14004206	hk2	hexokinase 2
ENSDARG00000034503	2.261103100242347e-16	5.567338300152267e-13	0.6082489350504545	2	48309600	48375342	per2	period circadian clock 2
ENSDARG00000102435	6.025610180317608e-17	1.9075360227976884e-13	1.0539265022132713					
ENSDARG00000101482	9.742460938723084e-17	2.6986616800262944e-13	0.9350743176658163					
ENSDARG00000034503	2.261103100242347e-16	5.567338300152267e-13	0.6082489350504545					

# Look up genes in databases

GENE

**noxo1a**

**ID** ZDB-GENE-030131-9700

**Name** *NADPH oxidase organizer 1a*

**Symbol** *noxo1a* Nomenclature History

**Previous Names** *noxo1*, *cb18* (1), *sb:cb18*, *SNX28b* (1), *wu:fd09d09*, *zgc:152911* (1)

**Type** [protein\\_coding\\_gene](#)

**Location** Chr: 3 [Mapping Details/Browsers](#)

**Description** Predicted to have phosphatidylinositol-3-phosphate binding activity and superoxide-generating NADPH oxidase activator activity. Predicted to be involved in superoxide metabolic process. Predicted to localize to NADPH oxidase complex and cytoplasm. Is expressed in EVL; periderm; and pharynx. Orthologous to human **NOXO1** (NADPH oxidase organizer 1).

**Genome Resources** [Alliance](#) (1), [Gene:572245](#) (1), [Ensembl\(GRCz11\):ENSDARG00000041294](#) (3)

**Note** None

**Comparative Information** 

# Look up genes in databases

GENE

 noxo

## NOXO1

ID		
Name		
Symbol		
Previous N	Species	<i>Homo sapiens</i>
Type	Symbol	NOXO1
Location	Name	NADPH oxidase organizer 1
Description	Synonyms	MGC20258 NADPH oxidase regulatory protein <a href="#">▼ Show All 12</a>
Genome Re		
Note	Biotype	protein coding gene
Comparativ	Automated Description <a href="#">?</a>	Enables enzyme binding activity. Involved in extracellular matrix disassembly. Part of NADPH oxidase complex.
Information		
RGD Description	This gene encodes an NADPH oxidase (NOX) organizer, which positively regulates NOX1 and NOX3. The protein contains a PX domain and two SH3 domains. Alternatively spliced transcript variants encoding multiple isoforms have been observed for this gene. [provided by RefSeq, Jun 2012]	
Cross References	<a href="#">ENSEMBL:ENSG00000196408</a> <a href="#">NCBI_Gene:124056</a> <a href="#">▼ Show All 4</a>	
Additional Information	<a href="#">Literature</a>	

# Look up genes in databases

GENE

noxo

GENE

NOXO1

ID	Species
Name	NOXO1
Symbol	NOXO1
Previous Name	
Type	Gene
Location	
Description	Synonyms
Genome Reference	
Note	
Comparative Information	
RGD Description	
Cross References	

**Summaries for NOXO1 Gene**

**Entrez Gene Summary for NOXO1 Gene**

This gene encodes an NADPH oxidase (NOX) organizer, which positively regulates NOX1 and NOX3. The protein contains a PX domain and two SH3 domains. Alternatively spliced transcript variants encoding multiple isoforms have been observed for this gene. [provided by RefSeq, Jun 2012]

**GeneCards Summary for NOXO1 Gene**

NOXO1 (NADPH Oxidase Organizer 1) is a Protein Coding gene. Diseases associated with NOXO1 include [Lung Mucoepidermoid Carcinoma](#) and [Phagocyte Bactericidal Dysfunction](#). Among its related pathways are [Signaling by Rho GTPases](#) and [Disease](#). Gene Ontology (GO) annotations related to this gene include *identical protein binding* and *phospholipid binding*. An important paralog of this gene is [SH3PXD2A](#).

**UniProtKB/Swiss-Prot Summary for NOXO1 Gene**

Constitutively potentiates the superoxide-generating activity of NOX1 and NOX3 and is required for the biogenesis of otoconia/otolith, which are crystalline structures of the inner ear involved in the perception of gravity. Isoform 3 is more potent than isoform 1 in activating NOX3. Together with NOXA1, may also substitute to NCF1/p47phox and NCF2/p67phox in supporting the phagocyte NOX2/gp91phox superoxide-generating activity. ( [NOXO1\\_HUMAN,Q8NFA2](#) )

**Gene Wiki entry for NOXO1 Gene**

**Additional gene information for NOXO1 Gene**

[HGNC \(19404\)](#) [NCBI Entrez Gene \(124056\)](#) [Ensembl \(ENSG00000196408\)](#) [OMIM® \(611256\)](#) [UniProtKB/Swiss-Prot \(Q8NFA2\)](#)  
[Open Targets Platform \(ENSG00000196408\)](#)

Alliance of Genome Resources

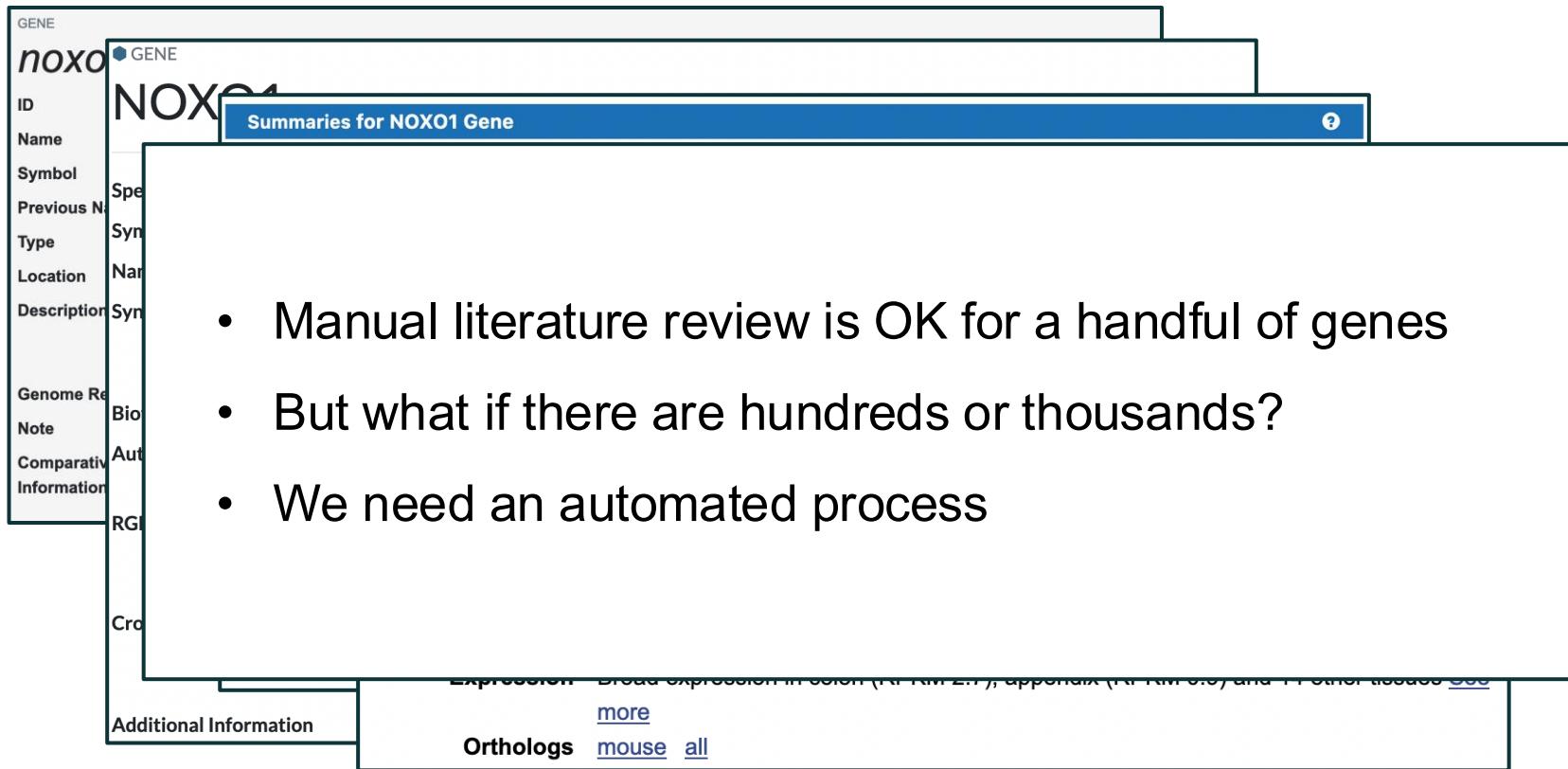
Additional Information

Literature

# Look up genes in databases

Summaries for NOXO1 Gene	
Entrez Gene Summary	<a href="#">View</a>
Species	<b>Official Symbol</b> NOXO1 provided by <a href="#">HGNC</a>
Symbol	<b>Official Full Name</b> NADPH oxidase organizer 1 provided by <a href="#">HGNC</a>
Previous Name	<b>Primary source</b> <a href="#">HGNC:HGNC:19404</a>
Type	<b>See related</b> <a href="#">Ensembl:ENSG00000196408</a> <a href="#">MIM:611256</a> ; <a href="#">AllianceGenome:HGNC:19404</a>
Location	<b>Gene type</b> protein coding
Description	<b>RefSeq status</b> REVIEWED
Genome Reference	<b>Organism</b> <a href="#">Homo sapiens</a>
Note	<b>Lineage</b> Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo
Comparative Information	<b>Also known as</b> SNX28; P41NOX; P41NOXA; P41NOXB; P41NOXC; SH3PXD5
RGD Description	<b>Summary</b> This gene encodes an NADPH oxidase (NOX) organizer, which positively regulates NOX1 and NOX3. The protein contains a PX domain and two SH3 domains. Alternatively spliced transcript variants encoding multiple isoforms have been observed for this gene. [provided by RefSeq, Jun 2012]
Cross References	<b>Expression</b> Broad expression in colon (RPKM 2.7), appendix (RPKM 0.9) and 14 other tissues <a href="#">See more</a>
	<b>Orthologs</b> <a href="#">mouse</a> <a href="#">all</a>
Additional Information	

# Look up genes in databases



The image shows a gene database interface for the gene NOXO1. The top navigation bar includes 'GENE', 'noxo1', 'ID', 'Name', 'Symbol', 'Previous N...', 'Type', 'Location', 'Description', 'Genome Re...', 'Note', 'Comparati...', 'Information', 'RGD', and 'Cross...'. The main content area features a blue header bar with 'Summaries for NOXO1 Gene' and a help icon. Below this, there is a large, empty white box. At the bottom, there are sections for 'Additional Information' (with a 'more' link), 'Orthologs' (with links for 'mouse' and 'all'), and 'Expression' (with a long, mostly illegible list of terms).

- Manual literature review is OK for a handful of genes
- But what if there are hundreds or thousands?
- We need an automated process

# Functional enrichment analysis

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- **Functional enrichment analysis** (or over-representation) systematically relates your data to existing knowledge
- Can help you to:
  - Gain biological insight
  - Generate new hypotheses
  - Validate your experiment

# Functional gene sets

- Existing knowledge is organised into **functional gene sets** in a standardised way, using data from previous experiments
- A functional gene set is a group of genes with a common biological relationship (e.g. annotated to same biological process or involved in same pathway)
- e.g. circadian rhythm:

Gene Product	Symbol	Qualifier	GO Term	Evidence	Reference	Assigned By	Name
UniProtKB:A0A024QZG3	ATF5	involved_in	GO:0007623    circadian rhythm	ECO:0000265 	GO_REF:0000107	Ensembl	BZIP domain-containing protein
UniProtKB:A0A024QZQ1	SIRT1	involved_in	GO:0007623    circadian rhythm	ECO:0000265 	GO_REF:0000107	Ensembl	Deacetylase sirtuin-type domain-containing protein
UniProtKB:A0A024R230	NTRK2	involved_in	GO:0007623    circadian rhythm	ECO:0000265 	GO_REF:0000107	Ensembl	Tyrosine-protein kinase receptor
UniProtKB:A0A024R241	NFIL3	involved_in	GO:0007623    circadian rhythm	ECO:0000256 	GO_REF:0000002	InterPro	Nuclear factor interleukin-3-regulated protein

# Functional annotation

- Functional annotation is created and maintained by many dedicated databases and projects, e.g.
  - Gene Ontology (GO)
  - Reactome
  - KEGG
  - TRANSFAC



# Gene Ontology

Current release 2026-01-23: 38,739 GO terms | 9,445,585 annotations  
1,723,113 gene products | 5,523 species (see statistics)

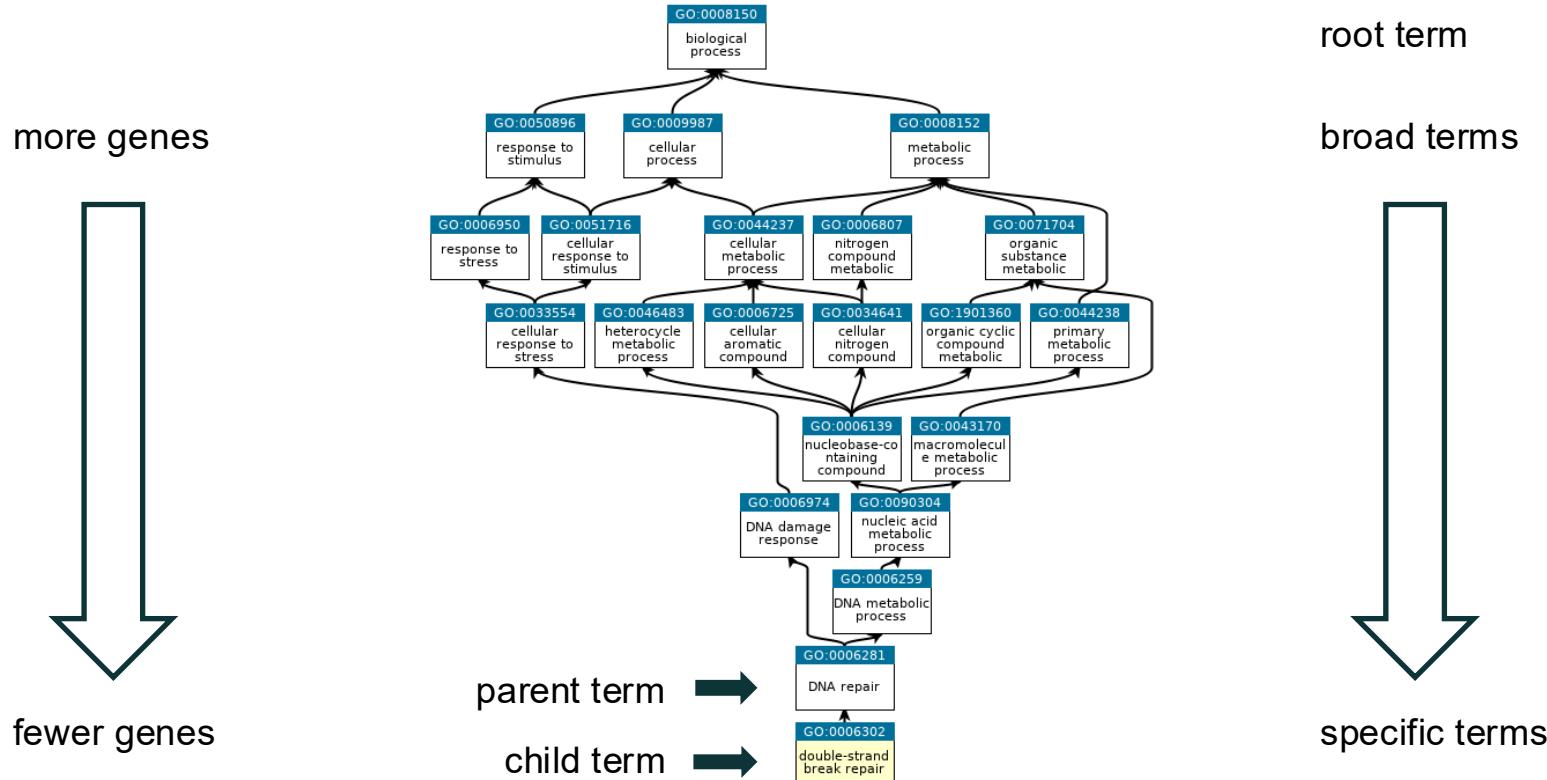
- GO is largest source of gene functional annotation
- Structured, controlled vocabulary of terms (and therefore gene sets)
- Manually annotated by a large consortium
- Data come from experimental and computational analyses

# GO ontologies

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- Actually three separate ontologies:
  - **Molecular Function** – molecular level activities performed by gene products, e.g. *transporter activity* (broad) or *Toll-like receptor binding* (specific)
  - **Cellular Component** – the cellular location where a function is performed, e.g. *ribosome*
  - **Biological Process** – larger processes accomplished by multiple molecular activities, e.g. *DNA repair* (broad) or *pyrimidine nucleobase biosynthetic process* (specific)
- Generally, in functional enrichment analysis, “biological process” is most useful

# GO hierarchy



# BRCA2 example

**Gene: BRCA2 ENSG00000139618**

**Description** BRCA2 DNA repair associated [Source:HGNC Symbol;Acc:HGNC:1101 ]

**Gene Synonyms** BRCC2, FACD, FAD, FAD1, FANCD, FANCD1, XRCC11

**Location** Chromosome 13: 32,315,086-32,400,268 forward strand. GRCh38:CM000675.2

**About this gene** This gene has 15 transcripts ([splice variants](#)), 173 [orthologues](#) and is associated with [120 phenotypes](#).

**Transcripts** [Show transcript table](#)

**GO: Molecular function** 

Show/hide columns (3 hidden)		Filter 	
Accession	Term	Evidence	Annotation source
GO:0002020 	protease binding	IPI	UniProt
GO:0003677 	DNA binding	IEA	UniProt
GO:0003697 	single-stranded DNA binding	IDA	UniProt
GO:0005515 	protein binding	IPI	IntAct
GO:0008022 	protein C-terminus binding	IDA	MGI
GO:0010484 	H3 histone acetyltransferase activity	IDA	UniProt
GO:0010485 	H4 histone acetyltransferase activity	IDA	UniProt
GO:0042802 	identical protein binding	IPI	IntAct
GO:0043015 	gamma-tubulin binding	IPI	UniProt

# BRCA2 example

## Gene: BRCA2 ENSG00000139618

Description	BRCA2 DNA repair associated [Source:HGNC Symbol;Acc:HGNC:1101]
Gene Synonyms	BRCC2, FACD, FAD, FAD1, FANCD, FANCD1, XRCC11
Location	<a href="#">Chromosome 13: 32,315,086-32,400,268</a> forward strand. GRCh38:CM000675.2
About this gene	This gene has 15 transcripts ( <a href="#">splice variants</a> ), 173 <a href="#">orthologues</a> and is associated with <a href="#">120 phenotypes</a> .
Transcripts	<a href="#">Show transcript table</a>

## GO: Molecular function

Show/hide columns (3 hidden)				Filter
Accession	Term	Evidence	Annotation source	
GO:0002020	protease binding	IPI	UniProt	
GO:0003677	DNA binding	IEA	UniProt	
GO:0003697	single-stranded DNA binding	IDA	UniProt	
GO:0005515	protein binding	IPI	IntAct	
GO:0008022	protein C-terminus binding	IDA	MGI	
GO:0010484	H3 histone acetyltransferase activity	IDA	UniProt	
GO:0010485	H4 histone acetyltransferase activity	IDA	UniProt	
GO:0042802	identical protein binding	IPI	IntAct	
GO:0043015	gamma-tubulin binding	IPI	UniProt	

## Gene: BRCA2 ENSG00000139618

Description	BRCA2 DNA repair associated [Source:HGNC Symbol;Acc:HGNC:1101]
Gene Synonyms	BRCC2, FACD, FAD, FAD1, FANCD, FANCD1, XRCC11
Location	<a href="#">Chromosome 13: 32,315,086-32,400,268</a> forward strand. GRCh38:CM000675.2
About this gene	This gene has 15 transcripts ( <a href="#">splice variants</a> ), 173 <a href="#">orthologues</a> and is associated with <a href="#">120 phenotypes</a> .
Transcripts	<a href="#">Show transcript table</a>

## GO: Cellular component

Accession	Term	Evidence	Annotation source
GO:0000152	nuclear ubiquitin ligase complex	IDA	ComplexPortal
GO:0000781	chromosome, telomeric region	IDA	BHF-UCL
GO:0000800	lateral element	IDA	MGI
GO:0005634	nucleus	IDA, IEA	UniProt
GO:0005654	nucleoplasm	IDA	HPA
GO:0005694	chromosome	IEA	Ensembl
GO:0005737	cytoplasm	IEA	UniProt
GO:0005813	centrosome	IDA	UniProt
GO:0005815	microtubule organizing center	IEA	UniProt
GO:0005829	cytosol	IDA	HPA
GO:0005856	cytoskeleton	IEA	UniProt
GO:0030141	secretory granule	IDA	UniProt
GO:0032991	protein-containing complex	IDA	MGI
GO:0033593	BRCA2-MAGE-D1 complex	IDA	UniProt
GO:1990391	DNA repair complex	IPI	ComplexPortal

# BRCA2 example

**Gene: BRCA2 ENSG00000139618**

**Description** BRCA2 DNA repair associated [Source:HGNC Symbol;Acc:HGNC:1101]

**Gene Synonyms** BRCC2, FACD, FAD, FAD1, FANCD, FANCD1, XRCC11

**Location** Chromosome 13: 32,315,086-32,400,268 forward strand. GRCh38:CM000675.2

**About this gene** This gene has 15 transcripts (splice variants), 173 orthologues and is associated with 120 phenotypes.

**Transcripts** [Show transcript table](#)

**GO: Biological process** [?](#)

Show	All	entries	Show/hide columns (3 hidden)	Filter
Accession	Term	Evidence	Annotation source	
GO:0000722	telomere maintenance via recombination	IEA	Ensembl	
GO:0000724	double-strand break repair via homologous recombination	IEA		
GO:0001556	oocyte maturation	IEA	Ensembl	
GO:0001833	inner cell mass cell proliferation	IEA	Ensembl	
GO:0006281	DNA repair	IEA		
GO:0006289	nucleotide-excision repair	IMP	UniProt	
GO:0006302	double-strand break repair	IMP	UniProt	
GO:0006310	DNA recombination	IEA	UniProt	
GO:0006355	regulation of DNA-templated transcription	IBA	GO_Central	
GO:0006974	cellular response to DNA damage stimulus	IEA	UniProt	
GO:0006978	DNA damage response, signal transduction by p53 class mediator resulting in transcription of p21 class mediator	IEA	Ensembl	
GO:0007049	cell cycle	IEA	UniProt	

**GO: Molecular function** [?](#)

Show/hide columns (3 hidden)	Filter		
Accession	Term	Evidence	Annotation source
GO:0002020	protease binding	IEA	Ensembl
GO:0003677	DNA binding	IEA	
GO:0003697	single-stranded DNA binding	IEA	
GO:0005515	protein binding	IEA	
GO:0008022	protein C-terminus binding	IEA	
GO:0010484	H3 histone acetyltransferase activity	IEA	
GO:0010485	H4 histone acetyltransferase activity	IEA	
GO:0042802	identical protein binding	IEA	
GO:0043015	gamma-tubulin binding	IEA	

00000139618

BRCA2 DNA repair associated [Source:HGNC Symbol;Acc:HGNC:1101]

BRCC2, FACD, FAD, FAD1, FANCD, FANCD1, XRCC11

Chromosome 13: 32,315,086-32,400,268 forward strand. GRCh38:CM000675.2

This gene has 15 transcripts (splice variants), 173 orthologues and is associated with 120 phenotypes.

[Show transcript table](#)

component [?](#)

Show/hide columns (3 hidden)	Filter		
Accession	Term	Evidence	Annotation source
GO:0000722	telomere maintenance via recombination	IEA	Ensembl
GO:0000724	double-strand break repair via homologous recombination	IEA	
GO:0001556	oocyte maturation	IEA	Ensembl
GO:0001833	inner cell mass cell proliferation	IEA	Ensembl
GO:0006281	DNA repair	IEA	
GO:0006289	nucleotide-excision repair	IMP	UniProt
GO:0006302	double-strand break repair	IMP	UniProt
GO:0006310	DNA recombination	IEA	UniProt
GO:0006355	regulation of DNA-templated transcription	IBA	GO_Central
GO:0006974	cellular response to DNA damage stimulus	IEA	UniProt
GO:0006978	DNA damage response, signal transduction by p53 class mediator resulting in transcription of p21 class mediator	IEA	Ensembl
GO:0007049	cell cycle	IEA	UniProt

# Functional enrichment analysis

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- How do we use all the existing annotation to interpret our gene list?
- Want to identify biological functions that are enriched in our gene list

# Testing for functional enrichment

20,000 genes assayed

Gene	adjp
ENSDARG00000041294	1.0867269119101765e-32
ENSDARG00000060498	1.2517176061993635e-21
ENSDARG00000031683	2.364463411626359e-21
ENSDARG00000077982	2.3548243274497384e-14
ENSDARG00000040480	5.7350346636925e-14
ENSDARG0000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075350227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG00000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11
ENSDARG000000204748	3.388766458381055e-11
ENSDARG000000102808	3.652027442583773e-11
ENSDARG00000059294	3.686666857888031e-11
ENSDARG000000858731	7.374599119303466e-11
ENSDARG00000030896	1.0690865837043268e-10
ENSDARG000000117300	2.836221657423099e-10
ENSDARG000000102758	7.675777258160306e-10
ENSDARG00000004754	1.3907777984676073e-9
ENSDARG000000099960	2.0021043563850804e-9
ENSDARG000000056615	2.032136501204873e-9
ENSDARG000000073820	3.749723988428957e-9
ENSDARG000000003570	4.027615069700461e-9
ENSDARG000000077799	7.49209763602349e-9
ENSDARG000000058386	7.75409869629005e-9
ENSDARG000000094678	7.81554132440011e-9
ENSDARG000000076914	2.810237900118536e-8
ENSDARG000000037421	2.990985153534242e-8
ENSDARG000000014340	3.505316564907291e-8
ENSDARG000000104773	4.1069391754303735e-8
ENSDARG000000105749	4.1069391754303735e-8
ENSDARG000000018491	4.275633454932327e-8
ENSDARG000000109648	4.9384742480335793e-8
ENSDARG000000010231	5.822859593920341e-8
ENSDARG000000039142	7.665381608460826e-8
ENSDARG000000104672	1.2504769486538527e-7
ENSDARG000000054196	1.356347303797386e-7
ENSDARG0000000090548	1.8758159693858449e-7
ENSDARG000000104919	2.2367046789114162e-7
ENSDARG000000062788	3.7107672341197336e-7
ENSDARG0000000079227	4.32879091031615297e-7
ENSDARG000000051888	4.6980261652096274e-7
ENSDARG0000000077169	4.7089485706046475e-7
ENSDARG000000056196	4.7089485706046475e-7
ENSDARG000000018283	4.917667288531457e-7
ENSDARG000000105731	5.385812733974355e-7
ENSDARG000000004954	5.526970490118169e-7
ENSDARG000000025903	7.245067792685657e-7
ENSDARG000000025094	7.999474861543812e-7
ENSDARG000000100504	8.037125469772779e-7

Adjusted  
p-value  
< 0.05



500 significantly DE genes

Gene	adjp
ENSDARG00000041294	1.0867269119101765e-32
ENSDARG00000060498	1.2517176061993635e-21
ENSDARG00000031683	2.364463411626359e-21
ENSDARG00000077982	2.9548243274497384e-14
ENSDARG000000040480	5.735034663692255e-14
ENSDARG0000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075360227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG00000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11

# Testing for functional enrichment

20,000 genes assayed

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ENSDARG00000041294	1.0867269119101765e-32
ENSDARG00000060498	1.2517176061993635e-21
ENSDARG00000031683	2.364463411626339e-21
ENSDARG00000077982	2.9548243274497384e-14
ENSDARG00000004080	5.735034663692255e-14
ENSDARG00000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075360227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG000000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11
ENSDARG0000000204748	3.38876645838105e-11
ENSDARG000000102808	3.65202742583773e-11
ENSDARG00000059294	3.68666685788831e-11
ENSDARG00000085871	7.374599119303466e-11
ENSDARG00000030896	1.0690865837043268e-10
ENSDARG000000117300	3.836221657423099e-10
ENSDARG000000102758	7.67577725816030e-10
ENSDARG00000004754	1.3907777984676073e-9
ENSDARG000000099960	2.0021043563850804e-9
ENSDARG000000056615	2.032136501204873e-9
ENSDARG000000073820	3.749723988428957e-9
ENSDARG00000003570	4.827615069700461e-9
ENSDARG000000077799	7.49209763602349e-9
ENSDARG000000058386	7.75409869629005e-9
ENSDARG000000094678	7.81554132440011e-9
ENSDARG000000076914	2.810237900118536e-8
ENSDARG000000037421	2.990985153534242e-8
ENSDARG000000014340	3.505316564907291e-8
ENSDARG000000104773	4.1069391754303735e-8
ENSDARG000000105749	4.1069391754303735e-8
ENSDARG000000018491	4.275633454932327e-8
ENSDARG000000109648	4.9384742480335793e-8
ENSDARG000000010231	5.822859593920341e-8
ENSDARG000000039142	7.665381608460826e-8
ENSDARG000000104672	1.2504769486538527e-7
ENSDARG000000054196	1.356347303797386e-7
ENSDARG000000090548	1.8758159693858449e-7
ENSDARG000000104919	2.2367046789114162e-7
ENSDARG000000062788	3.7107672341197336e-7
ENSDARG000000079227	4.3287909103165297e-7
ENSDARG000000051888	4.6980261652096274e-7
ENSDARG000000077169	4.7089485706046475e-7
ENSDARG000000056196	4.7089485706046475e-7
ENSDARG000000018283	4.917667288531457e-7
ENSDARG000000105731	5.885812733974355e-7
ENSDARG000000004954	5.526970430118169e-7
ENSDARG000000025903	7.245067792685657e-7
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ENSDARG000000070480	5.735034663692255e-14
ENSDARG00000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075360227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG000000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11

200 genes annotated  
to DNA repair

$$200/500 = 40\%$$

(300 not annotated to  
DNA repair)

2000 genes annotated to  
function (e.g. DNA repair)

$$2000/20000 = 10\%$$

(18,000 not annotated to  
DNA repair)

# Testing for functional enrichment

20,000 genes assayed

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ENSDARG00000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075350227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG000000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11
ENSDARG0000000204748	3.3887645838105e-11
ENSDARG000000102808	3.65027442583773e-11
ENSDARG00000059294	3.68666857888831e-11
ENSDARG00000085871	7.374599119303466e-11
ENSDARG00000030896	1.0690865837043268e-10
ENSDARG000000117300	3.836221657423099e-10
ENSDARG000000102758	7.67577725816030e-10
ENSDARG00000004754	1.3907777984676073e-9
ENSDARG000000099960	2.0021043563850804e-9
ENSDARG000000056615	2.032130501204873e-9
ENSDARG000000073820	3.749723988428957e-9
ENSDARG00000003570	4.027615069700461e-9
ENSDARG000000077799	7.49209763602349e-9
ENSDARG000000053836	7.75409869629005e-9
ENSDARG000000094678	7.8155413244001e-9
ENSDARG000000076914	2.810237900118536e-8
ENSDARG000000037421	2.99098513533424e-8
ENSDARG000000014340	3.505316564907291e-8
ENSDARG000000104773	4.1069391754303735e-8
ENSDARG000000105749	4.1069391754303735e-8
ENSDARG000000018491	4.275633454933237e-8
ENSDARG000000109648	4.9384742480335793e-8
ENSDARG000000010231	5.822859593920341e-8
ENSDARG000000039142	7.665381608460826e-8
ENSDARG000000104672	1.250476948653827e-7
ENSDARG000000054196	1.356347303797386e-7
ENSDARG000000090548	1.8758159693858449e-7
ENSDARG000000104919	2.2367646789114162e-7
ENSDARG000000062788	3.7107672341197336e-7
ENSDARG000000079227	4.822859593920341e-7
ENSDARG000000051888	6.9802616520996274e-7
ENSDARG0000000077169	4.7089485706046475e-7
ENSDARG000000056196	4.7089485706046475e-7
ENSDARG000000018283	4.917667288531457e-7
ENSDARG000000105731	5.885812733974355e-7
ENSDARG000000004954	5.526970430118169e-7
ENSDARG000000025903	7.245067792685657e-7
ENSDARG000000025094	7.999474861543812e-7
ENSDARG000000100504	8.037125469772779e-7

Adjusted  
p-value  
< 0.05

500 significantly DE genes

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ENSDARG00000077982	2.9548243274497384e-14
ENSDARG00000070480	5.735034663692255e-14
ENSDARG00000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075360227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG00000013670	2.3318547986985375e-11
ENSDARG00000039490	3.047413661053603e-11

2000 genes annotated to  
function (e.g. DNA repair)

2000/20000 = 10%

(18,000 not annotated to  
DNA repair)

200 genes annotated  
to DNA repair

200/500 = 40%

(300 not annotated to  
DNA repair)

Is seeing 200 DNA repair  
genes significantly differentially  
expressed more than we would  
expect by chance?

# Testing for functional enrichment

20,000 genes assayed

Gene	adjp
ENSDARG00000041294	1.0867269119101765e-32
ENSDARG00000060498	1.2517176061993635e-21
ENSDARG00000031683	2.364463411626359e-21
ENSDARG00000077982	2.9548243274497384e-14
ENSDARG000000040480	5.735034663692255e-14
ENSDARG00000007769	1.5678213864306653e-13
ENSDARG000000102435	1.9075350227976884e-13
ENSDARG000000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG000000013670	2.33185479869985375e-11
ENSDARG00000039490	3.047413661053603e-11
ENSDARG0000000204748	3.3887645838105e-11
ENSDARG000000102808	3.65027442583773e-11
ENSDARG00000059294	3.68666857888831e-11
ENSDARG00000005871	7.374599119303466e-11
ENSDARG00000030896	1.0690865837043268e-10
ENSDARG000000117300	3.836221657423099e-10
ENSDARG000000102758	7.67577725816030e-10
ENSDARG00000004754	1.3907777984676073e-9
ENSDARG000000099960	2.0021043563850804e-9
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ENSDARG000000105749	4.1069391754303735e-8
ENSDARG000000018491	4.2756334594332327e-8
ENSDARG000000109648	4.9384742480335793e-8
ENSDARG000000010231	5.822859593020341e-8
ENSDARG000000039142	7.665381608460826e-8
ENSDARG000000104672	1.250476948653827e-7
ENSDARG000000051416	1.356347303797386e-7
ENSDARG000000050548	1.8758159693858449e-7
ENSDARG000000104919	2.2367047891114162e-7
ENSDARG00000002788	3.7107672341197336e-7
ENSDARG000000079227	4.2879095103165297e-7
ENSDARG000000051888	4.6980261652096274e-7
ENSDARG000000077169	4.7089485706046475e-7
ENSDARG000000056196	4.7089485706046475e-7
ENSDARG000000018283	4.917667288531457e-7
ENSDARG000000105731	5.885812733974355e-7
ENSDARG00000004954	5.526970430118169e-7
ENSDARG000000025903	7.245067732685657e-7
ENSDARG000000025094	7.999474861543812e-7
ENSDARG000000105004	8.0371254697727799e-7

Adjusted  
p-value  
< 0.05

500 significantly DE genes

Gene	adjp
ENSDARG00000041294	1.0867269119101765e-32
ENSDARG00000060498	1.2517176061993635e-21
ENSDARG00000031683	2.364463411626359e-21
ENSDARG00000077982	2.9548243274497384e-14
ENSDARG00000070480	5.735034663692255e-14
ENSDARG00000007769	1.5678213864306653e-13
ENSDARG00000102435	1.9075360227976884e-13
ENSDARG00000101482	2.6986616800262944e-13
ENSDARG00000034503	5.567338300152267e-13
ENSDARG00000013670	2.33185479869985375e-11
ENSDARG00000039490	3.047413661053603e-11

200 genes annotated  
to DNA repair

$$200/500 = 40\%$$

(300 not annotated to  
DNA repair)

2000 genes annotated  
function (e.g. DNA

$$2000/20000 = 10\%$$

(18,000 not annotated  
DNA repair)

	DE	Not DE	Total
Annotated to DNA repair	200	1800	2000
Not annotated to DNA repair	300	17700	18000
<b>Total</b>	<b>500</b>	<b>19500</b>	<b>20000</b>

# Hypergeometric test

	DE	Not DE	Total
Annotated to DNA repair	200	1800	2000
Not annotated to DNA repair	300	17700	18000
<b>Total</b>	<b>500</b>	<b>19500</b>	<b>20000</b>

```
> m <- 20000 # Total genes
> n <- 500 # Number of DE genes
> mt <- 2000 # Number of annotated genes
> nt <- 200 # Number of annotated DE genes
> phyper(nt - 1, mt, m - mt, n, lower.tail=FALSE)
[1] 1.65531e-72
```

Use the hypergeometric test to calculate the probability of having 200 or more DE annotated genes when 2000 of the 20,000 total genes are annotated

$$P(\sigma_t \geq n_t) = \sum_{k=n_t}^{\min(m_t, n)} \frac{\binom{m_t}{k} \binom{m-m_t}{n-k}}{\binom{m}{n}}$$

# Multiple testing correction

---

- In reality, won't just be doing one test
- Want to test all (or a lot) of the GO terms and other functional gene sets
- Leads to problem of **multiple testing**
- If you test 10,000 GO terms with a significance threshold of  $< 0.05$  then you expect 500 terms to be significant simply by chance
- Need to correct for multiple testing:
  - Bonferroni
  - Benjamini–Hochberg

# Bonferroni correction

- Bonferroni is easiest to understand and most conservative
- Simply multiply all p-values by the number of tests (i.e. functional gene sets)
- Get adjusted p-values

GO	pval	adjp
GO:0022008	5.947e-7	5.947e-6
GO:0008038	8.705e-7	8.705e-6
GO:0097367	0.000001	0.000010
GO:0043168	0.000002	0.000020
GO:0010975	0.004917	0.049172
GO:0036211	0.005152	0.051521
GO:0021631	0.020739	0.207394
GO:0065009	0.272362	1.000000
GO:0099545	0.290182	1.000000
GO:1905245	0.496883	1.000000

# Benjamini–Hochberg correction

---

- Benjamini–Hochberg is less conservative and assumes that all tests are statistically independent
- Not true – many functional gene sets overlap:
  - e.g. GO terms are hierarchical so a term's annotations are a subset of their parental annotations
  - e.g. similar pathways can appear in KEGG and WikiPathways
  - e.g. some genes are co-expressed
- Nevertheless, BH is widely and successfully used
- Although Wijesooriya *et al.* (2022) found that 43% of papers surveyed failed to do multiple testing correction:  
[doi.org/10.1371/journal.pcbi.1009935](https://doi.org/10.1371/journal.pcbi.1009935)

# Background gene set

---

- Important to choose appropriate background gene set
- Wijesooriya *et al.* (2022) found that only 4% of papers used an appropriate background (although most failed to specify what background was used):  
[doi.org/10.1371/journal.pcbi.1009935](https://doi.org/10.1371/journal.pcbi.1009935)
- Best to choose all genes that could have been captured in your experiment
- Examples:
  - All genes
  - All genes with non-zero total read count in DESeq2
  - All genes that pass DESeq2 independent filtering
  - All genes expressed in a particular tissue
  - All genes with annotations

# Other methods

---

- Functional enrichment analysis (or over-representation analysis) is just one method
- Other methods and tests are available, e.g.
  - GSEA (gene set enrichment analysis)
  - Binomial test
- Concentrating on functional enrichment analysis because most widely used and most tools available

# Advantages of functional enrichment analysis

---

- Improves statistical power as you effectively sum up counts from the multiple genes in a functional gene set
- Improves statistical power as there are usually fewer functional annotations than genes, so less multiple testing correction is needed
- Results are easier to interpret because they are familiar concepts like “DNA repair” rather than obscure gene names
- Diverse data (e.g. RNA-seq, proteomics) can be integrated because they map to common terms/pathways
- Results may be more comparable to related data because results are projected to a smaller set of functional annotations

# Disadvantages of functional enrichment analysis

---

- Terms or pathways with few genes are unlikely to ever be enriched
- Hypergeometric test is more likely to identify larger functional gene sets (e.g. pathways with many genes) as significant
- Genes with multiple functions can lead to enrichment of multiple terms/pathways, some of which aren't relevant
- Databases are (obviously) biased towards genes with annotation so unannotated genes (e.g. many non-coding RNA genes) are invisible to functional enrichment analysis

# Recommendations based on disadvantages

---

- For human RNA-seq data, consider excluding functional gene sets with < 10 genes and > 500 genes
- Former are unlikely to ever be significant and latter are too likely to be significant and will often be better represented by other more specific terms/pathways
- Always think about your own experiment:
  - e.g. is apoptosis enrichment expected or a symptom of a problem during sample preparation

# Quiz!

---

- Quiz on Mentimeter ([www.menti.com](http://www.menti.com))

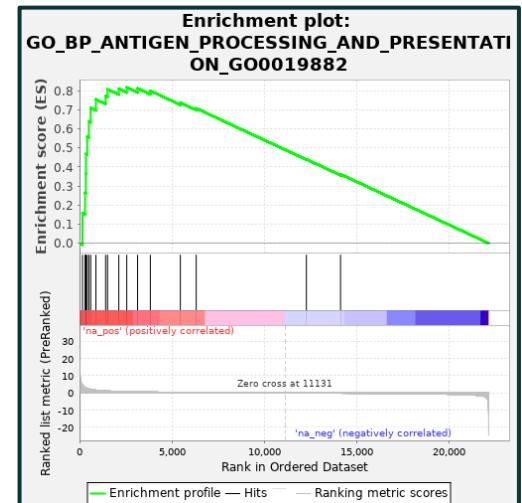
# Functional enrichment tools

- Many, many functional enrichment analysis tools exist
- Many are created, published and then never updated
- Best to choose a well used tool
- Using g:Profiler because:
  - Consistently and regularly updated over many years
  - Easy to use
  - Free
  - Well documented
  - Has advanced features, like simultaneous analysis of multiple lists
  - Has web interface but also an API with supported R and Python packages
  - Covers nearly 800 species/strains/varieties



# Other functional enrichment tools

- Other tools are available (and good):
  - Enrichr ([maayanlab.cloud/Enrichr/](http://maayanlab.cloud/Enrichr/)):
    - Web-based
    - Similar to g:Profiler
    - Only human, mouse, fly, yeast, worm and zebrafish
  - GSEA ([www.gsea-msigdb.org/gsea/](http://www.gsea-msigdb.org/gsea/)):
    - Desktop software
    - Implements GSEA method
    - Works on whole genome ranked gene lists
    - Looks for gene sets enriched at top or bottom of your ranked list
    - p-values computed by permutating ranked lists



# g:Profiler

---

- g:Profiler uses Ensembl as its primary data source (specifically, BioMart)
- Tracks Ensembl release schedule (every three or four months) but with delay of weeks or months
- Since July last year, g:Profiler had been using Ensembl 113, which came out in October 2024
- Recommend using Ensembl IDs as input, but not essential

g:Profiler

News Archives Beta API R client FAQ Docs Contact Cite g:Profiler Services using g:P GMT Helper

≡

**g:GOSt**  
Functional profiling

**g:Convert**  
Gene ID conversion

**g:Orth**  
Orthology search

**g:SNPense**  
SNP id to gene name

Query Upload query Upload bed file

Input is whitespace-separated list of genes [?](#)

Advanced options

Data sources

Bring your data (Custom GMT)

Run query random example mixed query example

## Options

Organism: [?](#)

Homo sapiens (Human)

Highlight driver terms in GO [?](#)

Ordered query [?](#)

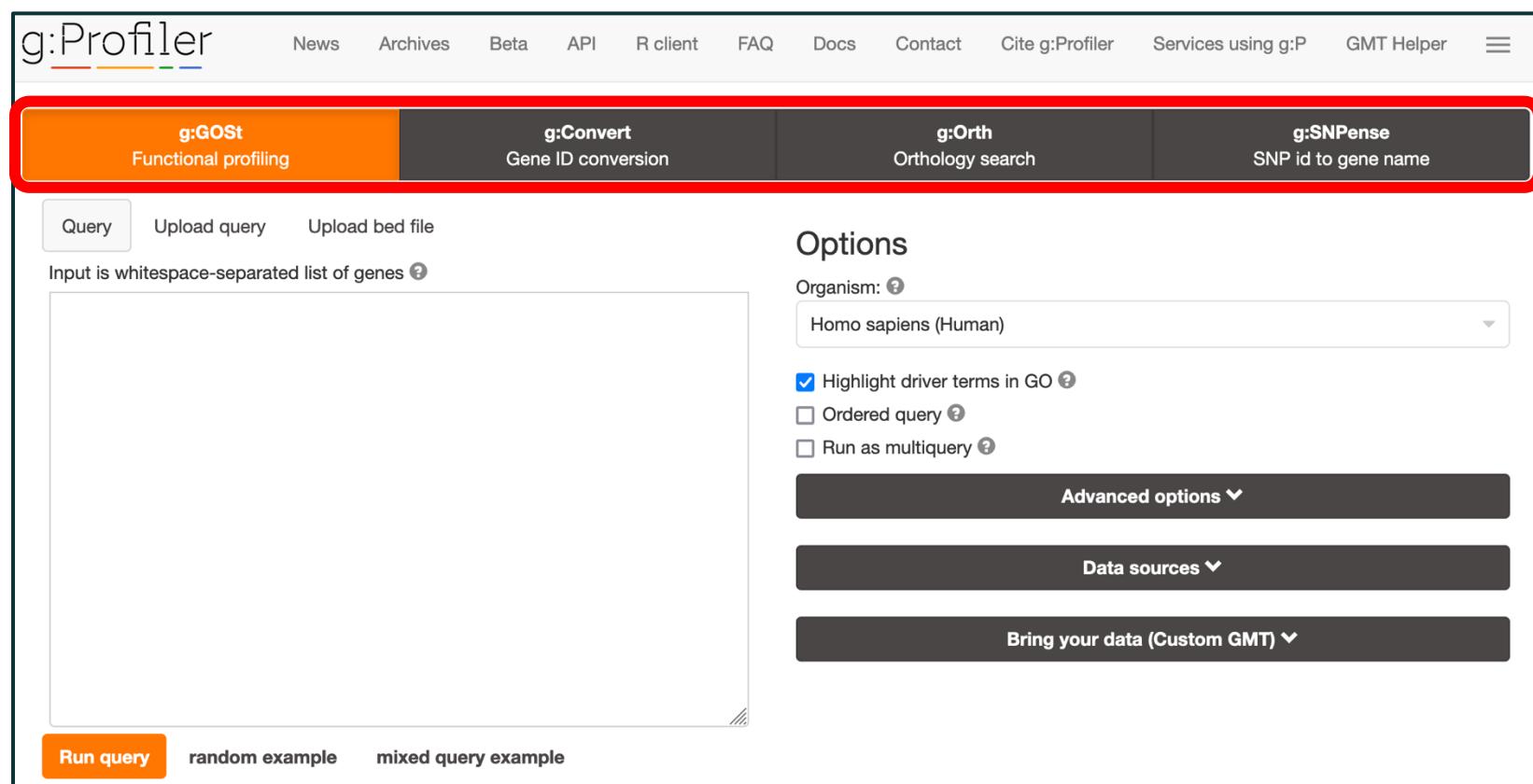
Run as multiquery [?](#)

**Advanced options**

**Data sources**

**Bring your data (Custom GMT)**

# g:Profiler – four tools



The screenshot shows the g:Profiler homepage with a red box highlighting the first tool in the navigation bar: g:GOSt. The navigation bar also includes g:Convert, g:Orth, and g:SNPense. Below the navigation bar, there are three input options: Query, Upload query, and Upload bed file. A text input field is labeled "Input is whitespace-separated list of genes" with a question mark icon. To the right, there is a "Options" section with a dropdown for "Organism" set to "Homo sapiens (Human)". Under "Options", there are three checkboxes: "Highlight driver terms in GO" (checked), "Ordered query" (unchecked), and "Run as multiquery" (unchecked). Below these are three buttons: "Advanced options", "Data sources", and "Bring your data (Custom GMT)". At the bottom, there are three buttons: "Run query" (orange), "random example", and "mixed query example".

g:Profiler

News Archives Beta API R client FAQ Docs Contact Cite g:Profiler Services using g:P GMT Helper

g:GOSt  
Functional profiling

g:Convert  
Gene ID conversion

g:Orth  
Orthology search

g:SNPense  
SNP id to gene name

Query Upload query Upload bed file

Input is whitespace-separated list of genes ?

Organism: ?

Homo sapiens (Human)

Highlight driver terms in GO ?

Ordered query ?

Run as multiquery ?

Advanced options ▾

Data sources ▾

Bring your data (Custom GMT) ▾

Run query random example mixed query example

# g:Profiler – gene list

The screenshot shows the g:Profiler interface for a gene list analysis. The top navigation bar includes links for News, Archives, Beta, API, R client, FAQ, Docs, Contact, Cite g:Profiler, Services using g:P, and GMT Helper. Below the navigation is a horizontal menu bar with four tabs: g:GOSt (Functional profiling), g:Convert (Gene ID conversion), g:Orth (Orthology search), and g:SNPense (SNP id to gene name). The main content area has a red box around the 'Query' input field and its associated buttons: 'Upload query' and 'Upload bed file'. A text input placeholder says 'Input is whitespace-separated list of genes'. A callout box highlights the 'Options' section, which includes 'Organism' dropdown (set to 'Homo sapiens'), 'Highlight' checkbox (checked), 'Order by' dropdown (set to 'None'), and 'Run as multiquery' checkbox (unchecked). The callout box displays the following text: '101 identifiers recognised for human', '80 for mouse; 96 for zebrafish', and 'Advanced options', 'Data sources', 'Bring your data (Custom GMT)'. At the bottom are buttons for 'Run query' (orange), 'random example', and 'mixed query example'.

g:Profiler

News Archives Beta API R client FAQ Docs Contact Cite g:Profiler Services using g:P GMT Helper

g:GOSt Functional profiling g:Convert Gene ID conversion g:Orth Orthology search g:SNPense SNP id to gene name

Query Upload query Upload bed file

Input is whitespace-separated list of genes

101 identifiers recognised for human

80 for mouse; 96 for zebrafish

Advanced options

Data sources

Bring your data (Custom GMT)

Run query random example mixed query example

# g:Profiler – options

The screenshot shows the g:Profiler web interface. At the top, there is a navigation bar with links: News, Archives, Beta, API, R client, FAQ, Docs, Contact, Cite g:Profiler, Services using g:P, and GMT Helper. Below the navigation bar are four main tabs: g:GOSt (Functional profiling), g:Convert (Gene ID conversion), g:Orth (Orthology search), and g:SNPense (SNP id to gene name). The g:GOSt tab is highlighted with an orange background. Below the tabs, there are buttons for 'Query', 'Upload query', and 'Upload bed file'. A text input field is labeled 'Input is whitespace-separated list of genes' with a question mark icon. To the right of the input field, a large text box contains the message: '1055 species/strains/varieties in current release'. An arrow points from this text box to the 'Options' section, which is highlighted with a red box. The 'Options' section includes a dropdown for 'Organism' set to 'Homo sapiens (Human)', and three checkboxes: 'Highlight driver terms in GO' (checked), 'Ordered query' (unchecked), and 'Run as multiquery' (unchecked). Below these are three dark grey buttons with white text: 'Advanced options', 'Data sources', and 'Bring your data (Custom GMT)'. At the bottom of the interface are three buttons: 'Run query' (orange), 'random example', and 'mixed query example'.

g:Profiler

News Archives Beta API R client FAQ Docs Contact Cite g:Profiler Services using g:P GMT Helper

g:GOS<sup>t</sup>  
Functional profiling

g:Convert  
Gene ID conversion

g:Orth  
Orthology search

g:SNPense  
SNP id to gene name

Query Upload query Upload bed file

Input is whitespace-separated list of genes ?

1055 species/strains/varieties in current release

Organism: ?

Homo sapiens (Human)

Highlight driver terms in GO ?

Ordered query ?

Run as multiquery ?

Advanced options ▾

Data sources ▾

Bring your data (Custom GMT) ▾

Run query random example mixed query example

# g:Profiler – advanced options

Query    Upload query    Upload bed file

Input is whitespace-separated list of genes ?

g:SCS – “Set Counts and Sizes”

Accounts for hierarchical nature of GO

Less conservative than Bonferroni but more conservative than Benjamini-Hochberg

Organism: ?

Homo sapiens (Human)

Highlight driver terms in GO ?

Ordered query ?

Run as multiquery ?

**Advanced options ^**

All results ?

Measure underrepresentation ?

No evidence codes ?

Statistical domain scope ?

Only annotated genes

Significance threshold ?

g:SCS threshold

User threshold ?

0.05

Numeric IDs treated as ?

ENTREZGENE\_ACC

# g:Profiler – data sources

9 data sources

(or 11 if count GO as three separate sources)

All 9 not available for all species

→

**Data sources ▾**

**Gene Ontology**

- GO molecular function
- GO cellular component
- GO biological process
- No electronic GO annotations ?

**biological pathways**

- KEGG
- Reactome
- WikiPathways

**regulatory motifs in DNA**

- TRANSFAC
- miRTarBase

**protein databases**

- Human Protein Atlas
- CORUM

**Human phenotype ontology**

- HP

[name.gmt zip](#) [combined name.gmt](#)  
[ENSG.gmt zip](#) [combined ENSG.gmt](#)

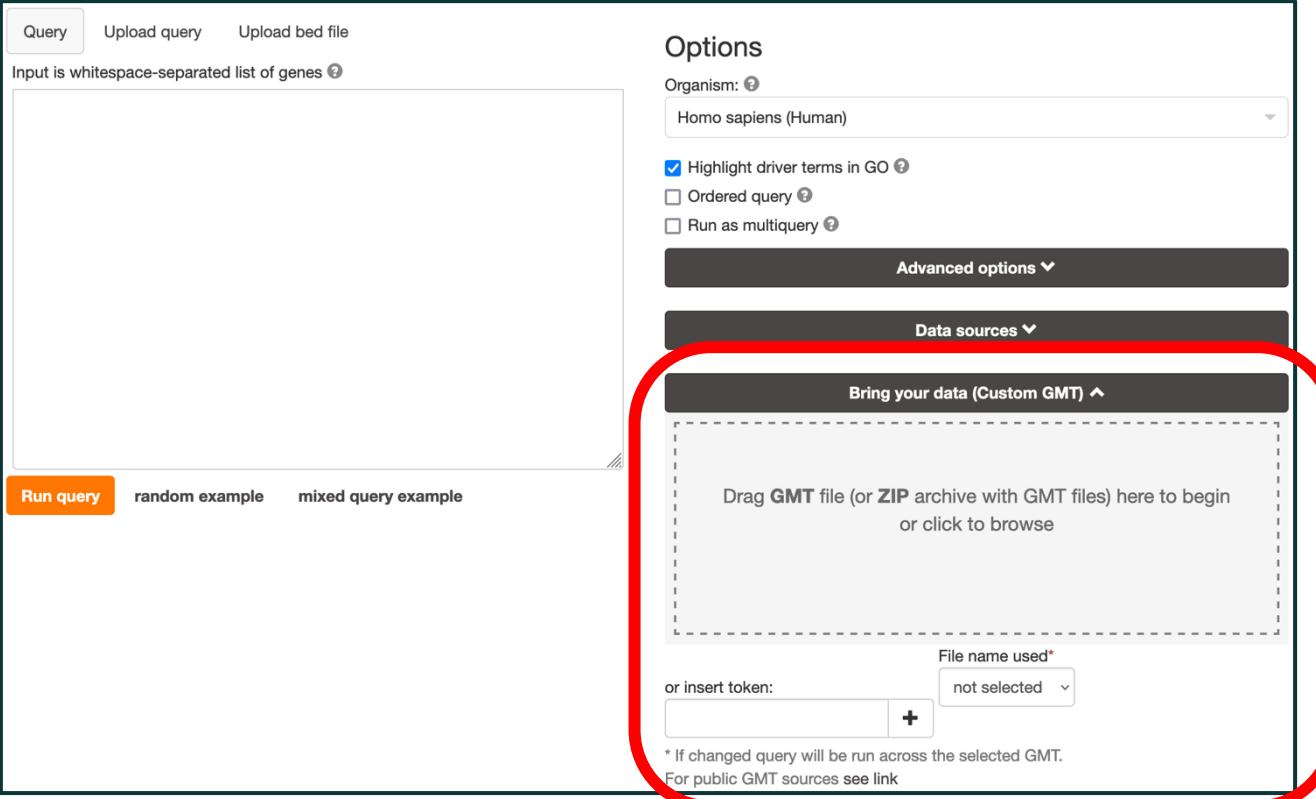
←

Can exclude GO IEA evidence term (inferred from electronic annotation)

But often as reliable as human annotation  
(Škunca et al. 2012)

Suggest running with and without if using human or model organisms

# g:Profiler – bring your data



The screenshot shows the g:Profiler web interface. At the top, there are three buttons: "Query" (highlighted in orange), "Upload query", and "Upload bed file". Below these is a text input field with placeholder text "Input is whitespace-separated list of genes ?". Underneath the input field are three buttons: "Run query" (orange), "random example", and "mixed query example". To the right of the input field is a "Options" section. It includes a dropdown for "Organism" set to "Homo sapiens (Human)", and three checkboxes: "Highlight driver terms in GO ?" (checked), "Ordered query ?" (unchecked), and "Run as multiquery ?" (unchecked). Below these are two buttons: "Advanced options ▾" and "Data sources ▾". The "Data sources" section is highlighted with a red box. It contains a "Bring your data (Custom GMT) ▾" section with a dashed box for dragging a GMT file or a ZIP archive. Below this is a "File name used\*" input field and a "not selected" dropdown. At the bottom of the "Bring your data" section is a note: "\* If changed query will be run across the selected GMT. For public GMT sources see link".

# g:Profiler – documentation

g:Profiler

News Archives Beta API R client FAQ **Docs** Contact Cite g:Profiler Services using g:P GMT Helper

g:GOST Functional profiling

g:Convert Gene ID conversion

g:Orth Orthology search

g:SNPense SNP id to gene name

## Welcome to g:Profiler

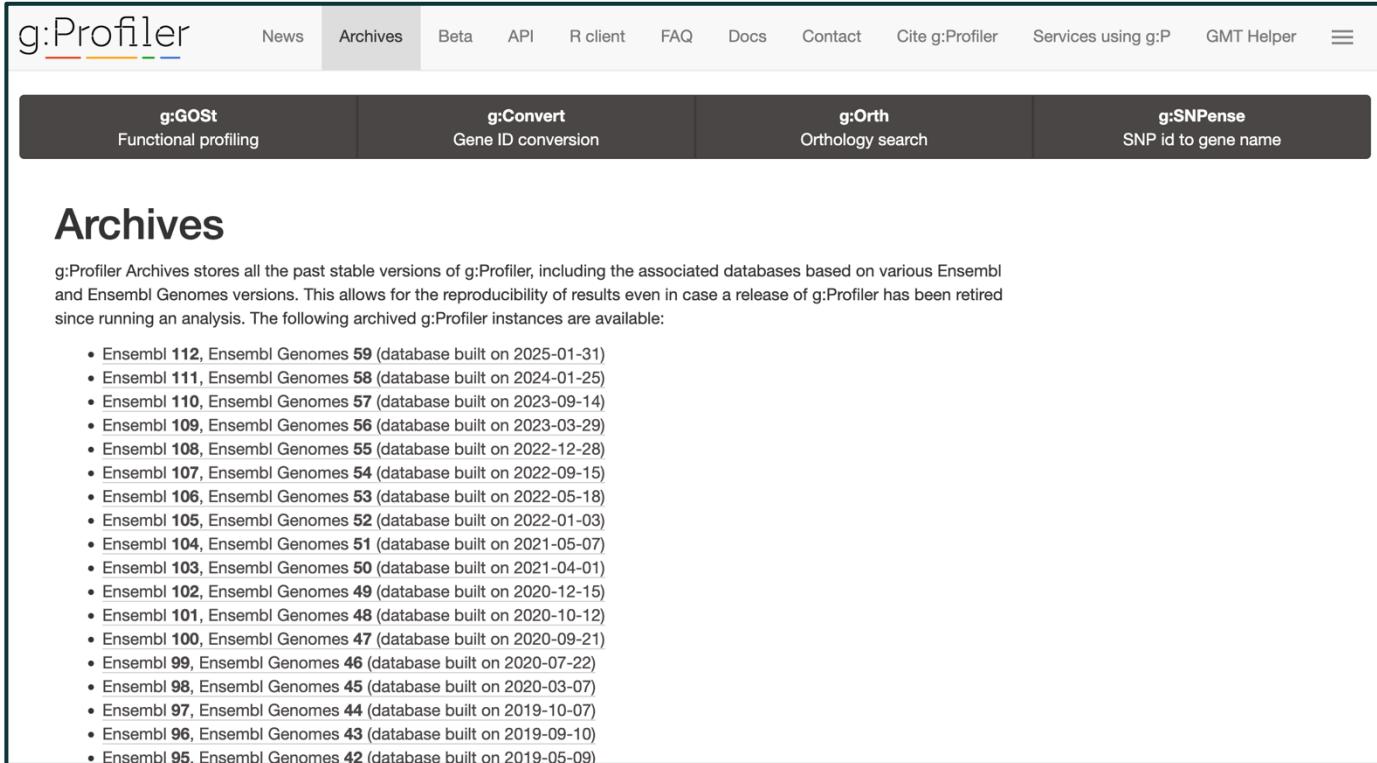
g:Profiler is a public web server for characterising and manipulating gene lists. g:Profiler has a simple user-friendly web interface with powerful visualisations and is currently available for 400+ species, including mammals, plants, fungi, insects from Ensembl and Ensembl Genomes. g:Profiler is updated approximately in every three months and follows quarterly releases of Ensembl databases. g:Profiler tool set consists of the following tools:

- **g:GOST**, the core of the g:Profiler, performs statistical enrichment analysis to provide interpretation to user-provided gene lists. The gene lists can be either flat or ordered gene lists. We accept majority of the identifier types, chromosomal regions and term IDs as input. We provide data from multiple sources of functional evidence, including Gene Ontology terms, biological pathways, regulatory motifs of transcription factors and microRNAs, human disease annotations and protein-protein interactions.
- **g:Convert** is a gene identifier conversion tool. It uses information in Ensembl databases to handle hundreds of types of IDs for genes, proteins, transcripts, microarray probesets, etc, for many species, experimental platforms and biological databases. g:Convert is flexible: it accepts a mixed list of IDs and recognises their types automatically. It can also serve as a service to get all genes belonging to a particular functional category.
- **g:Orth** is a tool for mapping homologous genes across related organisms based on Ensembl data. Given a selected target organism, g:Orth retrieves the genes of the target organism that are similar in sequence to the initial genes in the input.
- **g:SNPense** is a tool for mapping human single nucleotide polymorphisms (SNP) to gene names, chromosomal locations and variant consequence terms from Sequence Ontology.

Contents

- Welcome to g:Profiler
- About g:Profiler
- Publications and theses
- Funding
- Tech notes
- Support
- g:GOST
  - Using g:GOST
  - Highlighting
  - Examples
- g:Convert
  - Using g:Convert
  - Examples
- g:Orth
  - Using g:Orth
  - Examples
- g:SNPense

# g:Profiler – archives



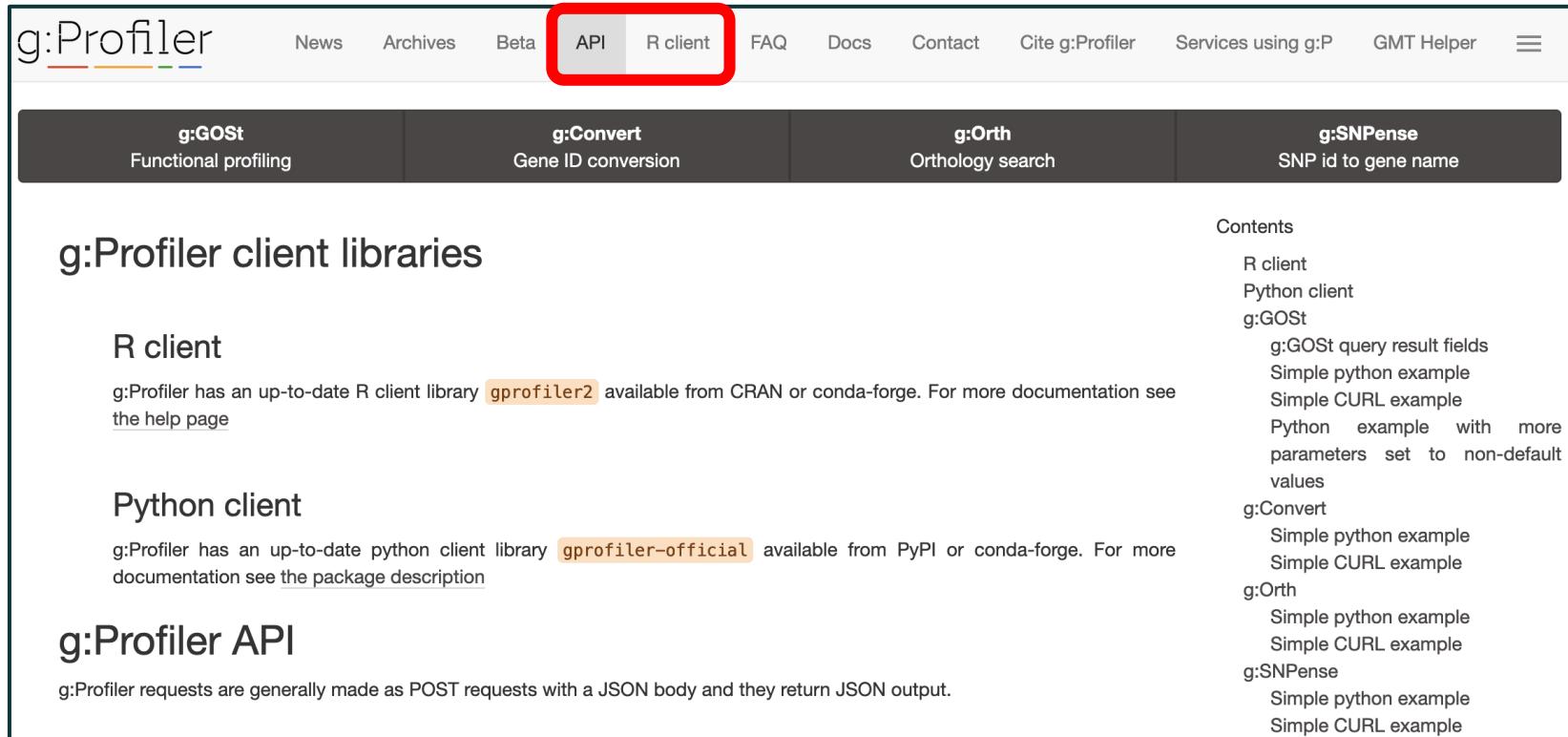
The screenshot shows the g:Profiler website with a teal header and a white main content area. The header includes a navigation bar with links: News, Archives (which is the active tab, highlighted in grey), Beta, API, R client, FAQ, Docs, Contact, Cite g:Profiler, Services using g:P, GMT Helper, and a menu icon. Below the header is a dark grey footer bar with four buttons: g:GOST (Functional profiling), g:Convert (Gene ID conversion), g:Orth (Orthology search), and g:SNPense (SNP Id to gene name).

## Archives

g:Profiler Archives stores all the past stable versions of g:Profiler, including the associated databases based on various Ensembl and Ensembl Genomes versions. This allows for the reproducibility of results even in case a release of g:Profiler has been retired since running an analysis. The following archived g:Profiler instances are available:

- [Ensembl 112](#), Ensembl Genomes [59](#) (database built on 2025-01-31)
- [Ensembl 111](#), Ensembl Genomes [58](#) (database built on 2024-01-25)
- [Ensembl 110](#), Ensembl Genomes [57](#) (database built on 2023-09-14)
- [Ensembl 109](#), Ensembl Genomes [56](#) (database built on 2023-03-29)
- [Ensembl 108](#), Ensembl Genomes [55](#) (database built on 2022-12-28)
- [Ensembl 107](#), Ensembl Genomes [54](#) (database built on 2022-09-15)
- [Ensembl 106](#), Ensembl Genomes [53](#) (database built on 2022-05-18)
- [Ensembl 105](#), Ensembl Genomes [52](#) (database built on 2022-01-03)
- [Ensembl 104](#), Ensembl Genomes [51](#) (database built on 2021-05-07)
- [Ensembl 103](#), Ensembl Genomes [50](#) (database built on 2021-04-01)
- [Ensembl 102](#), Ensembl Genomes [49](#) (database built on 2020-12-15)
- [Ensembl 101](#), Ensembl Genomes [48](#) (database built on 2020-10-12)
- [Ensembl 100](#), Ensembl Genomes [47](#) (database built on 2020-09-21)
- [Ensembl 99](#), Ensembl Genomes [46](#) (database built on 2020-07-22)
- [Ensembl 98](#), Ensembl Genomes [45](#) (database built on 2020-03-07)
- [Ensembl 97](#), Ensembl Genomes [44](#) (database built on 2019-10-07)
- [Ensembl 96](#), Ensembl Genomes [43](#) (database built on 2019-09-10)
- [Ensembl 95](#), Ensembl Genomes [42](#) (database built on 2019-05-09)

# g:Profiler – API and libraries



The screenshot shows the g:Profiler website with a red box highlighting the 'API' tab in the top navigation bar. The navigation bar also includes 'News', 'Archives', 'Beta', 'R client', 'FAQ', 'Docs', 'Contact', 'Cite g:Profiler', 'Services using g:P', 'GMT Helper', and a menu icon.

The main content area features four dark grey boxes with white text:

- g:GOSt**  
Functional profiling
- g:Convert**  
Gene ID conversion
- g:Orth**  
Orthology search
- g:SNPense**  
SNP id to gene name

**g:Profiler client libraries**

**R client**  
g:Profiler has an up-to-date R client library [gprofiler2](#) available from CRAN or conda-forge. For more documentation see [the help page](#)

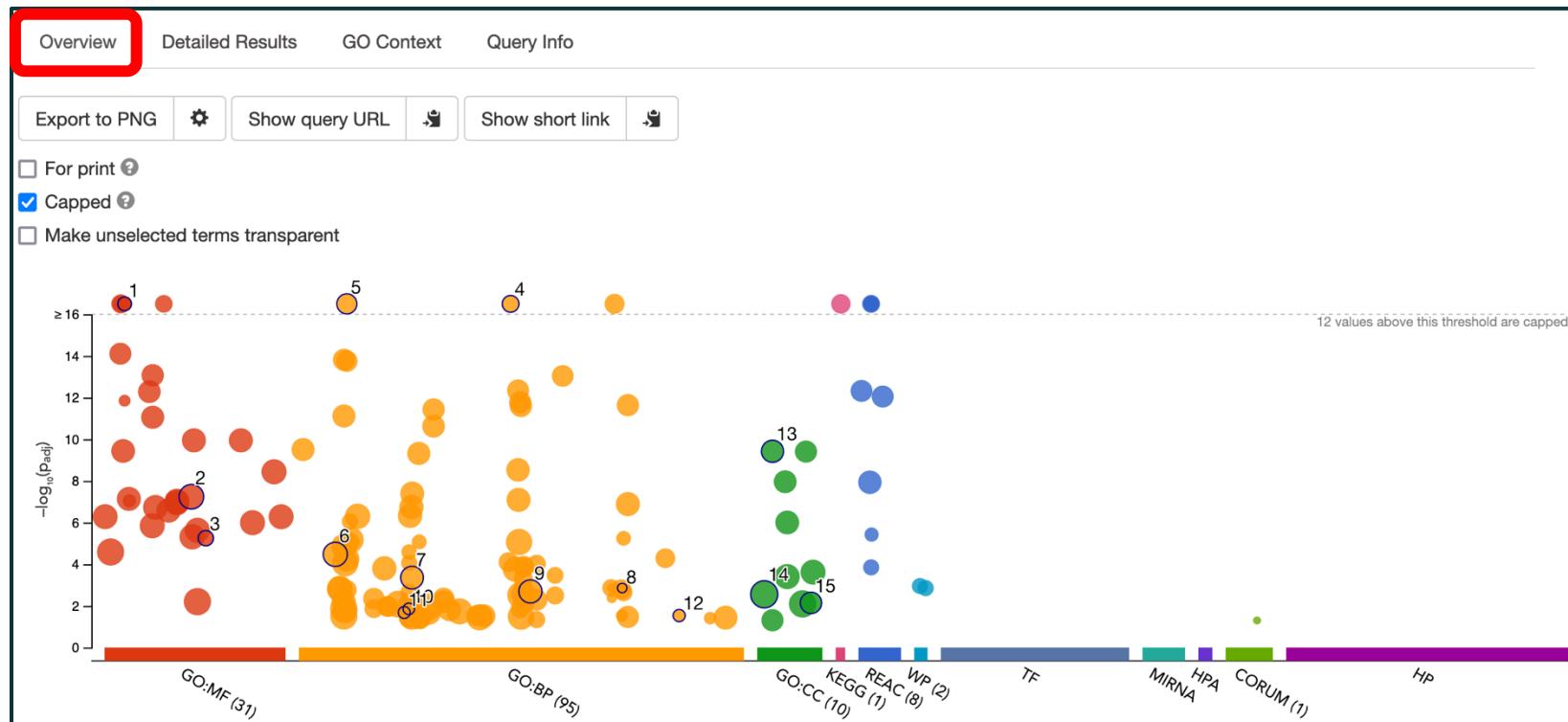
**Python client**  
g:Profiler has an up-to-date python client library [gprofiler-official](#) available from PyPI or conda-forge. For more documentation see [the package description](#)

**g:Profiler API**  
g:Profiler requests are generally made as POST requests with a JSON body and they return JSON output.

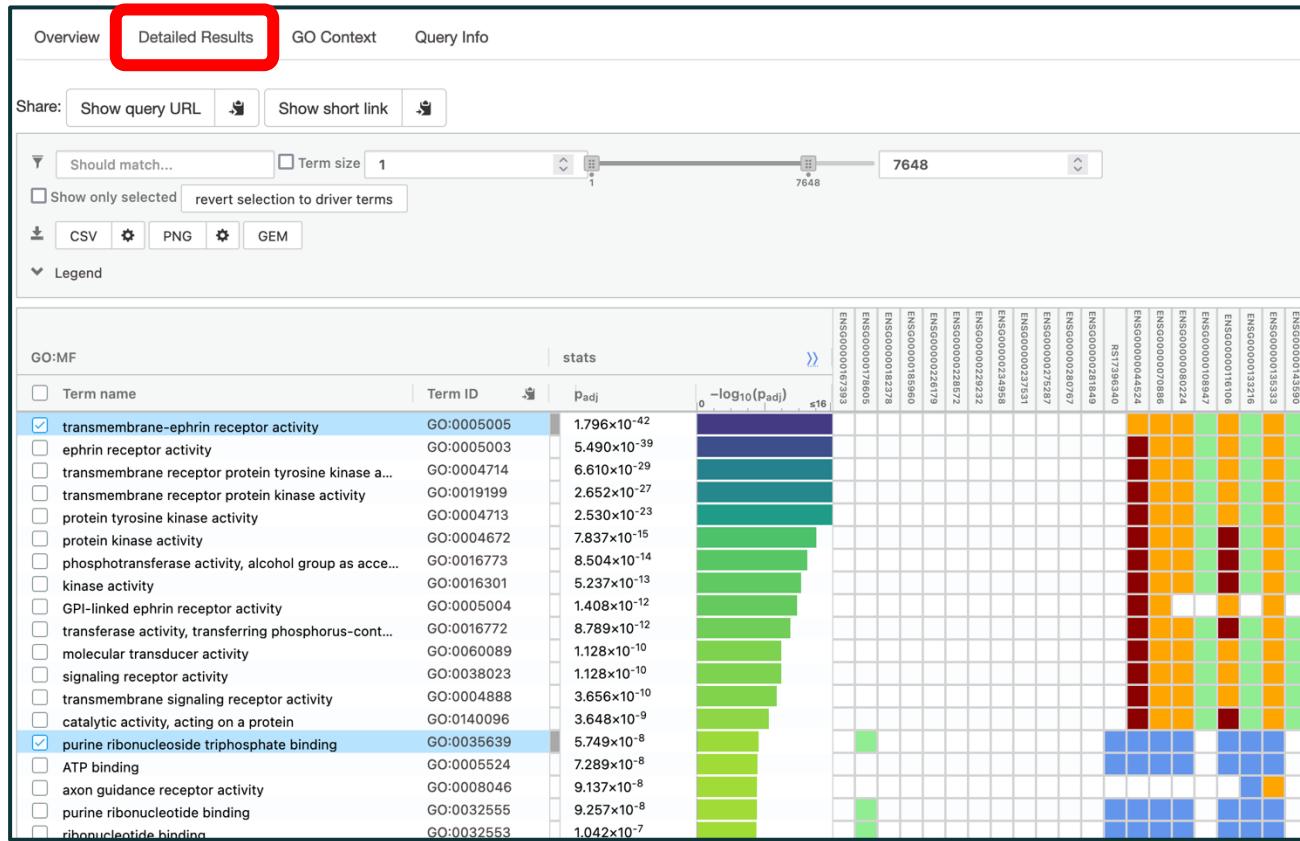
**Contents**

- R client
  - g:GOSt query result fields
  - Simple python example
  - Simple CURL example
  - Python example with more parameters set to non-default values
- g:Convert
  - Simple python example
  - Simple CURL example
- g:Orth
  - Simple python example
  - Simple CURL example
- g:SNPense
  - Simple python example
  - Simple CURL example

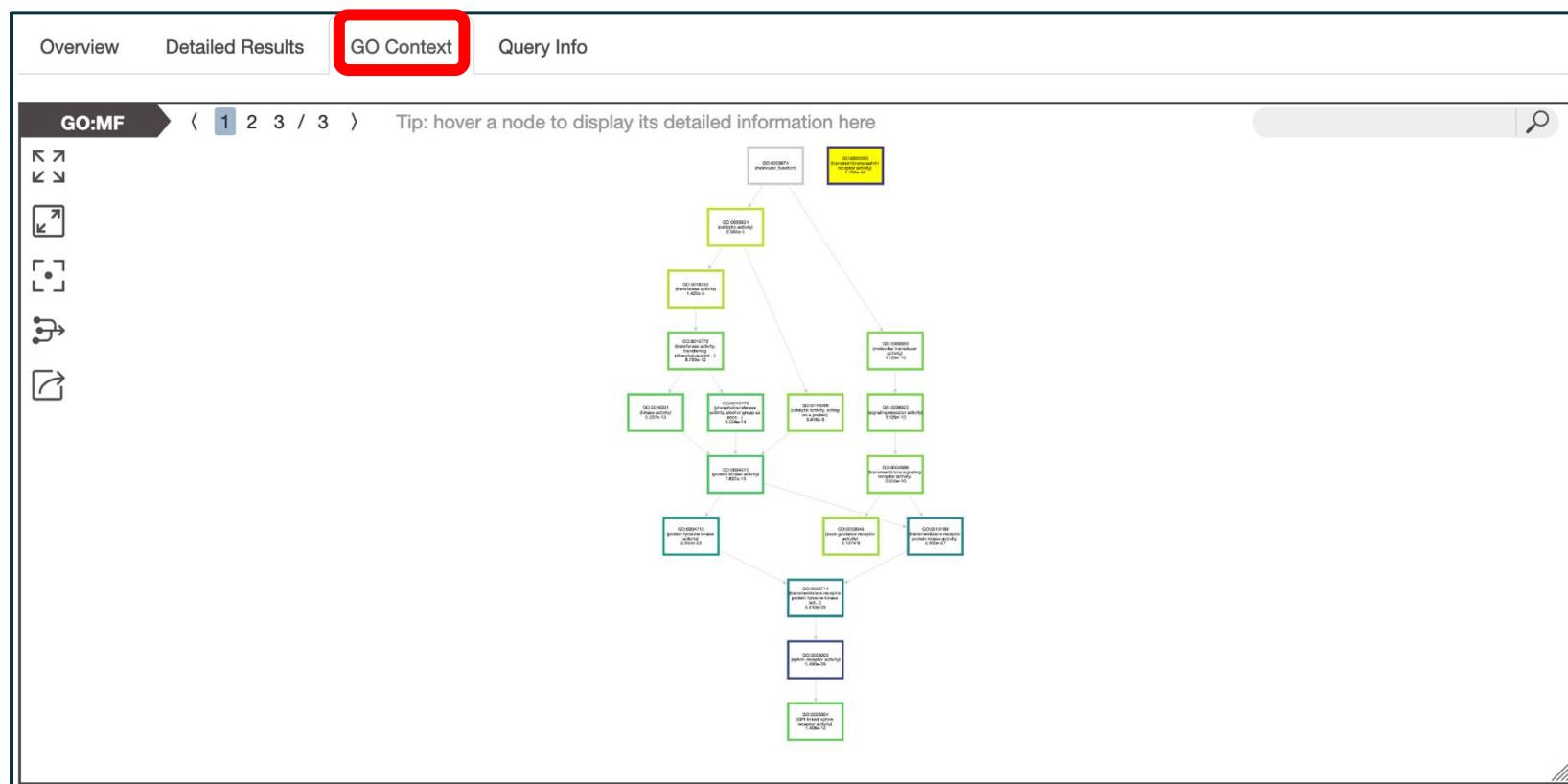
# g:Profiler – overview



# g:Profiler - detailed results



# g:Profiler – GO context



# g:Profiler – beta

The screenshot shows the g:Profiler beta website interface. The top navigation bar includes links for News, Archives, **Stable** (highlighted with a red box), API, R client, FAQ, Docs, Contact, Cite g:Profiler, Services using g:P, GMT Helper, and a menu icon. Below the navigation is a row of four main tools: g:GOSt (Functional profiling), g:Convert (Gene ID conversion), g:Orth (Orthology search), and g:SNPense (SNP id to gene name). The main content area features a query input field, a list of options, and three buttons at the bottom.

**g:Profiler <sup>β</sup>**

News Archives **Stable** API R client FAQ Docs Contact Cite g:Profiler Services using g:P GMT Helper ≡

**g:GOSt**  
Functional profiling

**g:Convert**  
Gene ID conversion

**g:Orth**  
Orthology search

**g:SNPense**  
SNP id to gene name

Query Upload query Upload bed file

Input is whitespace-separated list of genes ?

**Options**

Organism: ?

Homo sapiens (Human)

Highlight driver terms in GO ?

Ordered query ?

Run as multiquery ?

**Advanced options ▾**

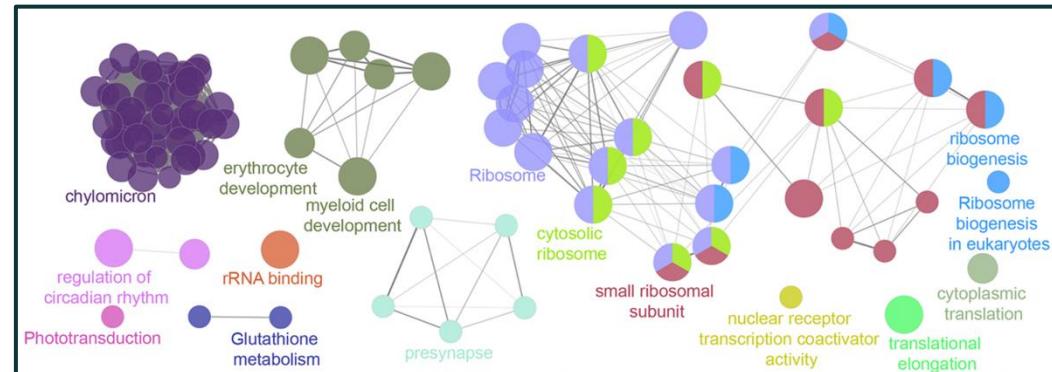
**Data sources ▾**

**Bring your data (Custom GMT) ▾**

**Run query** random example mixed query example

# Summarising functional enrichment

- Functional enrichment analysis (hopefully) summarises a gene list into something shorter and more comprehensible
- But what if the list of functional enrichments is also long and/or repetitive?
- The connected components functionality is an attempt to solve that problem
- Other methods:
  - Cytoscape / EnrichmentMap
  - Cytoscape / ClueGO
  - Revigo: <http://revigo.irb.hr/>



# g:Profiler live demo!

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- [biit.cs.ut.ee/gprofiler/](http://biit.cs.ut.ee/gprofiler/)

# Exercises (plus data and slides)

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- Exercises are available from:

[rnaseq2026.buschlab.org](http://rnaseq2026.buschlab.org)

- Plus data for exercises and these slides
- Everything also available on penelopeCloud